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REMARKS

The present application relates to hybrid maize plant and seed 34N16. Claims 1-42 are currently pending in the present application. Claims 9-11, 13-19, 22-24, 26-32, 34-40 have been canceled. Claims 43-54 have been added. Applicant respectfully requests consideration of the following remarks.

Detailed Action

A. Status of the Application

Applicant acknowledges the indefiniteness rejection and the art rejection of record of claims 10, 14, 18, 23, 27 and 31 have been overcome.

B. Specification

Applicant submits the Deposit section on page 50 has been amended in order to properly include both the hybrid maize plant 34N16 and the inbred parent plants GE568102 and GE534776 within the Deposit paragraph. The changes do not add new matter as there is literal support for the minor changes on pages 7 in the originally filed specification. The specification has now been amended to correct these minor changes.

In addition, Applicant respectfully submits that the actual ATCC deposit of the two inbred plants will be delayed until the receipt of notice that the application is otherwise in condition for allowance, in compliance under 37 C.F.R. §§ 1.801-1.809. Once such notice is received, an ATCC deposit will be made, and the specification will be amended to contain the accession number of the deposit, the date of the deposit, a description of the deposited biological material sufficient to specifically identify it and to permit examination and the name and address of the depository. The claims will also be amended to recite the ATCC deposit number. Applicant submits that at least 2,500 seeds of hybrid maize plant 34N16 and the inbred parents GE568102 and GE534776 will be deposited with the ATCC. Applicant further asserts that the deposits will be made without restriction

Rejections under 35 U.S.C. § 112, Second Paragraph

Claims 8, 11-19, 21, 24-32, 39, and 42 stand rejected under 35 U.S.C. § 112, second paragraph as being indefinite for failing to particularly point and distinctly claim the subject

matter which Applicant regards as the invention, as stated in the last Office Action for claims 5-8, 10-19, 21, and 23-32.

Applicant respectfully traverses this rejection. Applicant wishes to reiterate that it is well known in the art that the hybrid 34N16 will represent elite germplasm produced from the crossing of inbred parent lines GE568102 and GE534776 for character traits of major importance which will subsequently be used in a breeding population to further those elite traits. Applicant further asserts that it would be understood by one skilled in the art that the claimed maize plant or its parts contain at least 50% of the alleles inherited from the hybrid maize plant 34N16 having been deposited under ATCC Accession No. to be disclosed upon allowance of subject matter. Applicant further asserts, "[W]hen not defined by Applicant in the specification, the words of a claim must be given their plain meaning. In other words, they must be read as they would be interpreted by those of ordinary skill in the art", thereby alleviating this rejection. See *In re Sneed*, 710 F.2d 1544, 218 U.S.P.Q. 385 (Fed. Cir. 1983); See also MPEP § 2111.02. However, in order to expedite prosecution Applicant has canceled claims 9-11, 13-19, 22-24, 26-32, and 34-40, thereby alleviating this rejection to said claims.

The Examiner rejects claim 39 as indefinite for the recitation "34N16 maize plant ...deriving at least 50% of its alleles from 34N16" as confusing.

Applicant has now canceled claim 39, thus alleviating this rejection.

Claim 42 stands rejected as indefinite for failing to further limit claim 41.

Applicant has now amended claim 42 to read --A male sterile maize plant produced by the method of claim 41.--, as suggested by the Examiner. Applicant thanks the Examiner for the suggested language.

In light of the above amendments and remarks, Applicant respectfully requests reconsideration and withdrawal of the rejections under 35 U.S.C. § 112, second paragraph.

Rejections Under 35 U.S.C. § 112, First Paragraph

Claims 11, 15, 19, 24, 28, 32, 34 and 38-40 stand rejected under 35 U.S.C. § 112, first paragraph, as containing subject matter which was not described in the specification in such a way as to reasonably convey to one skilled in the relevant art that the inventor(s), at the time the application was filed, had possession of the claimed invention. The Examiner states that there is

no literal basis in the specification for the 50% allelic derivation language. Further the Examiner states that there is no basis for the double haploid method.

Applicant respectfully traverse this rejection. However, in an effort to expedite prosecution, Applicant has canceled claims 11, 15, 19, 24, 28, 32 and 38-40 and added new claims 43-54, alleviating this rejection. In addition, Applicant has now amended claims 12 and 25 to include --contains one or more transgenes which have been stably integrated therein, said transgenes selected from the group consisting of: a plant disease resistance gene, an insect resistance gene, a herbicide resistance gene, and a male sterility gene--, thereby limiting the claims to the types of transgenes that may be introduced and that are supported by the specification on pages 42-48, as suggested by the Examiner.

In addition, Applicant has canceled claims 34 and 40, thereby alleviating this rejection. Applicant respectfully asserts the following regarding double haploid breeding. The specification discusses multiple breeding techniques that may be used according to the invention. The specification at page 3 states [p]lant breeding techniques known in the art and used in a maize plant breeding program include, but are not limited to, recurrent selection backcrossing, pedigree breeding, restriction length polymorphism enhanced selection, genetic marker enhanced selection and transformation" (page 3, specification). Double haploid breeding is a technique long known and used in the art of plant breeding. Applicant is attaching herewith Wan *et al.*, "Efficient Production of Doubled Haploid Plants Through Colchicine Treatment of Anther-Derived Maize Callus", *Theoretical and Applied Genetics*, 77:889-892, 1989. This demonstrates haploid breeding is a long-known technique in the art of plant breeding and supports Applicant's assertion that producing double haploids is well known to one ordinarily skilled in the art. Further, it is axiomatic in patent law that a specification "need not teach, and preferably omits, what is well known in the art." See *Spectra-Physics, Inc. v. Coherent, Inc.*, 3 U.S.P.Q.2d 1737, 1743 (Fed. Cir. 1987). Double haploids are produced by the doubling of a set of chromosomes (1N) from a heterozygous plant to produce a completely homozygous individual. This is advantageous because the process can eliminate the generations of selfing needed to obtain a homozygous plant from a heterozygous source. Applicant therefore respectfully requests withdrawal of the above rejections.

Claim 33 stands rejected under 35 U.S.C. § 112, first paragraph as containing subject matter which was not described in the specification in such a way as to enable one skilled in the

art to which it pertains, or with which it is more nearly connected, to make and/or use the invention. The Examiner states the claim is drawn towards a method of making a hybrid plant designated 34N16 comprising crossing inbred maize plants GE568102 and GE534776, however the Examiner states the terms of this deposit are not known.

Applicant respectfully traverses this rejection. Applicant herein submits the Deposits section has been amended in order to properly include both the hybrid maize plant 34N16 and the inbred parents GE568102 and GE534776 within the Deposit paragraph on page 50. The changes do not add new matter as there is literal support for the minor changes on page 7 in the originally filed specification. The specification has now been amended to correct these minor changes. Applicant thanks the Examiner for pointing out this inadvertent mistake.

In addition Applicant submits that the actual ATCC deposit will be delayed until receipt of notice that the application is otherwise in condition for allowance. As provided in 37 C.F.R. §§ 1.801-1.809, Applicant wishes to reiterate they will refrain from deposit of hybrid 34N16 and inbred parents GE568102 and GE534776 until allowable subject matter is indicated. Once such notice is received, an ATCC deposit will be made, and the specification will be amended to contain the accession number of the deposit, the date of the deposit, description of the deposited biological materials sufficient to specifically identify and to permit examination and the name and address of the depository. The claims will also be amended to recite the proper ATCC deposit numbers. The Applicant provides assurance that:

- a) during the pendency of this application access to the invention will be afforded to the Commissioner upon request;
- b) all restrictions upon availability to the public will be irrevocably removed upon granting of the patent;
- c) the deposit will be maintained in a public depository for a period of thirty years, or five years after the last request for the enforceable life of the patent, whichever is longer;
- d) a test of the viability of the biological material at the time of deposit will be conducted (see 37 C.F.R. § 1.807); and
- e) the deposit will be replaced if it should ever become inviable.

Therefore, Applicant submits at least 2500 seeds of hybrid maize plant 34N16 and the inbred parents GE568102 and GE534776 will be deposited with the ATCC. In view of this assurance,

the rejection under 35 U.S.C. § 112, first paragraph, should be removed. (MPEP § 2411.02) Such action is respectfully requested.

Claims 8-19, 21-32 and 34-40 stand rejected under 35 U.S.C. § 112, first paragraph, as containing subject matter which is not described in the specification in such a way as to reasonably convey to one skilled in the art that the inventor(s), at the time the application was filed, had possession of the claimed invention, as stated in the last Office Action for claims 8-19 and 21-32. The Examiner stated that claims 12, 15, 25, 28 and dependents thereon are broadly drawn to any transgenic plant which contains any heterologous transgene of any sequence conferring any trait, and methods of using the transgenic plant. The Examiner further stated that claims 8, 16, 19, 21, 29, 32 and dependents thereon are broadly drawn to any "single gene conversion" plant comprising one or more traits introgressed into the claimed variety by backcrossing or other traditional means, and methods of using these plants.

Applicant respectfully traverses this rejection. However, in an effort to expedite prosecution, Applicant has canceled claims 9-11, 13-19, 22-24, 26-32, and 34-40 and amended claims 12 and 25 to include --contains one or more transgenes which have been stably integrated therein, said transgenes selected from the group consisting of: a plant disease resistance gene, an insect resistance gene, a herbicide resistance gene, and a male sterility gene--, thereby limiting the claims to the types of transgenes that may be introduced and that are supported by the specification as aforementioned.

Applicant wishes to reiterate that under the written description requirement, Applicant should be allowed to claim the progeny of a cross of maize plants crossed with 34N16 with phenotypic characteristics since distinguishing identifying characteristics in the chemical and biotechnological arts, dealing with DNA, are those such as: partial structure, physical and/chemical properties, functional characteristics, known or disclosed correlation between structure and function, method of making, and combinations of the above. In plants, these identifying characteristics are those detectable in the phenotype which are manifested through gene expression. Claims to a particular species of invention are adequately described in the disclosure of relevant identifying characteristics are present in the application. Again, one of ordinary skill in the art is reasonably apprised in knowing that a plant crossed with 34N16 will result in a plant having inherited half of the genetic material of 34N16. A further limitation set by Applicant is that the plants must be capable of expressing a combination of at least two of

these phenotypic characteristics of 34N16. Applicant respectfully submits the claims now come within the purview of the written description requirement and request reconsideration.

Claims 8, 12-19, 21, 25-32 and 34-40 stand rejected under 35 U.S.C. § 112, first paragraph, as containing subject matter which was not described in the specification in such a way as to enable one skilled in the art to which it pertains, or with which it is most nearly connected, to make and/or use the invention, for the reasons stated in the last Office Action for 8, 12-19, 21 and 25-32.

Applicant respectfully traverses this rejection. Applicant herein submits the Deposit section has been amended in order to properly include both hybrid maize plant 34N16 and the inbred parents GE568102 and GE534776 within the Deposit paragraph on page 50. The changes do not add new matter as there is literal support for the minor changes on page 7 in the originally filed specification. The Specification has now been amended to correct these minor changes. The Applicant further provides assurance that:

- a) during the pendency of this application access to the invention will be afforded to the Commissioner upon request;
- b) all restrictions upon availability to the public will be irrevocably removed upon granting of the patent;
- c) the deposit will be maintained in a public depository for a period of thirty years, or five years after the last request for the enforceable life of the patent, whichever is longer;
- d) a test of the viability of the biological material at the time of deposit will be conducted (see 37 C.F.R. § 1.807); and
- e) the deposit will be replaced if it should ever become inviable.

Therefore, Applicant submits at least 2500 seeds of hybrid maize plant 34N16 and the inbred parents GE568102 and GE534776 have been deposited with the ATCC. In view of this assurance, the rejection under 35 U.S.C. § 112, first paragraph, should be removed. (MPEP § 2411.02). In addition, Applicant submits a patent application "need not teach, and preferably omits, what is well known in the art." *Hybritech Inc. v. Monoclonal Antibodies Inc.*, 802 F.2d 1367, 231 U.S.P.Q. 81 (Fed. Cir. 1986); MPEP § 601. One of ordinary skill in the art of plant breeding would know how to evaluate the traits of two plant varieties to determine if there is no significant difference between the two traits expressed by those varieties. In addition, in an effort to expedite prosecution claims 9-11, 13-19, 22-24, 26-32, and 34-40 have been canceled.

For the reason aforementioned, it is respectfully submitted that Applicants' claims are sufficiently enabled by the specification.

In light of the above amendments and remarks, Applicant respectfully requests reconsideration and withdrawal of the rejections to claims 8-19 and 21-40 under 35 U.S.C. § 112, first paragraph.

Summary

Applicant acknowledges that claims 1-7, 20, and 41 are allowed.

Applicant acknowledges that claims 1-10, 12-14, 16-18, 20-23, 25-27 and 29-31 are deemed free of the prior art. The Examiner further states the prior art fails to teach or fairly suggest plants which derive 50% or more of their alleles from the exemplified hybrid. This clearly indicates that hybrid maize plant 34N16 as a whole is considered to be distinguishable from the prior art for the purposes of novelty and non-obviousness. Therefore, Applicant respectfully submits that the deposit of the representative seed of 34N16 and inbred parents GE568102 and GE534776 should satisfy the description requirement. In light of the above, Applicant respectfully submits that the rejections under 35 U.S.C. § 112, first paragraph as improper and requests reconsideration and withdrawal of these rejections.

Conclusion

In conclusion, Applicant submits in light of the above amendments and remarks, the claims as amended are in a condition for allowance, and reconsideration is respectfully requested.

No additional fees or extensions of time are believed to be due in connection with this amendment; however, consider this a request for any extension inadvertently omitted, and charge any additional fees to Deposit Account No. 26-0084.

Reconsideration and allowance is respectfully requested.

Respectfully submitted,



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Efficient production of doubled haploid plants through colchicine treatment of anther-derived maize callus

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Summary. A chromosome doubling technique, involving colchicine treatment of an embryogenic, haploid callus line of maize (*Zea mays* L., derived through anther culture), was evaluated. Two colchicine levels (0.025% and 0.05%) and three treatment durations (24, 48, and 72 h) were used and compared to untreated controls. Chromosome counts and seed recovery from regenerated plants were determined. No doubled haploid plants were regenerated from calli without colchicine treatment. After treatment with colchicine for 24 h, the callus tissue regenerated about 50% doubled haploid plants. All of the plants regenerated from the calli treated with colchicine for 72 h were doubled haploids, except for a few tetraploid plants. No significant difference in chromosome doubling was observed between the two colchicine levels. Most of the doubled haploid plants produced viable pollen and a total of 107 of 136 doubled haploid plants produced from 1 to 256 seeds. Less extensive studies with two other genotypes gave similar results. These results demonstrate that colchicine treatment of haploid callus tissue can be a very effective and relatively easy method of obtaining a high frequency of doubled haploid plants through anther culture.

Key words: *Zea mays* – Anther culture – Embryogenic haploid callus – Chromosome doubling

Introduction

The success of producing haploid plants in maize through anther culture makes it possible to generate

inbred lines through chromosome doubling (Kuo et al. 1986). However, the application of anther culture to plant breeding is largely dependent on the production of large numbers of haploid plants and the high frequency of induction of chromosome doubling. In maize, anther-derived lines have been developed and used commercially (Wu et al. 1983). However, the frequency of chromosome doubling of anther-derived haploid plants either spontaneously or through colchicine treatment has been undesirably low (Ku et al. 1978; Nitsch et al. 1982; Miao et al. 1978). Ku et al. (1978) and Nitsch et al. (1982) observed only 6.3% and 4.5% spontaneously doubled haploids among plants regenerated from cultured maize anthers, respectively. Miao et al. (1978) treated anther-derived plantlets and obtained only one plant which set seeds from the 24 plants that survived.

With many plant species, chromosome doubling can be achieved by the use of an antimutagenic agent treatment of anther-derived, haploid plantlets. Since antimutagenic agents such as colchicine usually induce chromosome doubling in only some cells due to the asynchrony of cell divisions, chimeric plants are common after colchicine treatment. For plant species which produce bisexual flowers and tillers or branches, chimeras are acceptable since some tillers or branches may develop from the chromosome-doubled cells. In contrast, maize plants usually do not produce tillers and cell lines which give rise to the tassel and ear are already determined in the mature seed (Coe and Neuffer 1978). Colchicine treatment of maize seedlings or plantlets may double the chromosome number in the tassel or ear, but often not in both, which will make self-pollination impossible. These reasons may explain why the efficiency of inducing doubled haploid plants in maize is very low by colchicine treatment of regenerated haploid plantlets (Miao et al. 1978).

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Since somatic embryos from tissue cultures may develop from one or a few cells, it may be possible to induce chromosome doubling in embryogenic haploid callus and then induce plant regeneration from this tissue (Genovesi and Collins 1982). The use of a long term haploid culture system capable of plant regeneration may make the chromosome doubling technique effective as proposed by Tsay et al. (1986). This paper reports the recovery of doubled haploid plants with high frequency through colchicine treatment of embryogenic haploid callus initiated from maize anther culture.

Materials and methods

Establishment of callus cultures. F1 plants of a maize hybrid, H99 × Frl6, were grown in the field in 1987. Tassel collection to anther plating were carried out by previously described methods (Petolino and Thompson 1987). Petri dishes containing anthers were placed in plastic boxes covered with aluminum foil at 28 °C. About 1 month later, embryo-like structures began to appear from responding anthers. The embryo-like structures were removed from the anthers and were transferred to a callus induction medium. The callus induction medium consisted of macronutrients and vitamins of N6 medium (Chu et al. 1975), micronutrients of B5 medium (Gamborg et al. 1968) with 2,4-D (0.45 µM), dicamba (11.3 µM), myo-inositol (0.55 mM), L-proline (25.0 mM), enzymatic casein hydrolysate (0.1 g/l), sucrose (87.6 mM), Na₂EDTA (110.55 µM) and FeSO₄ · 7 H₂O (100.2 µM). Callus lines, each of which was derived from a single embryo-like structure, were maintained in the callus induction medium through subcultures by selective transfer of the embryogenic calli at 4-week intervals. One highly regenerable callus line was used 6 months after culture initiation.

Colchicine treatment. Colchicine was dissolved in water to make a stock solution which was filter-sterilized and then added to liquid D medium (Duncan et al. 1985) to the required final concentrations and stored in the dark. About 20 ml of the medium was placed in Petri dishes (100 × 25 mm) and a filter paper disc supported by a stainless steel screen, which were autoclaved previously, was saturated with the liquid medium. Embryogenic calli, 20 days after subculture, were cut into 0.5–1.0 mm pieces and were plated on the moist filter paper and incubated in the dark at 28 °C. Following treatment, the calli were placed on a stainless steel screen and were rinsed twice in liquid D medium without colchicine. Two colchicine levels (0.025% and 0.05%) and three treatment durations were used and compared with untreated control.

Plant regeneration from treated calli. Colchicine treated calli were subcultured two times with an interval of 10 days on agar-solidified D medium. For plant regeneration, calli were transferred to H medium (Duncan et al. 1985) with 3.5 mg/l 6-benzyladenine for 3 days. The calli were then cultured in H medium until some regenerated plantlets grew to 3–4 cm long, which occurred within about 20 days. The regenerated plantlets were transferred to H medium minus RT vitamins and glucose in culture tubes for further growth. After 7–10 days, they were transplanted to soil in 11.5-cm pots and grown for another 10–15 days (or even longer depending on the growth of each plant). Finally, the plants were transplanted to 27.5-cm pots in the greenhouse and at least two root tips were collected from each plant for mitotic examination.

Plants with pollen and silks were self-pollinated on successive days. The dates of the first day of pollen shed and the first day of silk emergence were recorded for 33 representative doubled haploid plants. Seeds were harvested 40–45 days after pollination.

Determination of ploidy level. The root tips were cold-treated in ice water for 24 h and fixed in 3:1, 95% ethanol:glacial acetic acid for 24 h and then stored in 70% ethanol. For mitotic examination, the root tips from each plant were placed in a small vial with 1% acetocarmine and heated to the boiling point several times. The meristematic region was excised and squashed in one drop of 45% glacial acetic acid on a slide. At least two root tips from each regenerated plant were examined to determine the ploidy level.

Results

All 24 plants regenerated from the untreated calli contained the haploid number of ten chromosomes (Table 1, Fig. 1a). Of 96 plants regenerated from calli treated for 24 h with either 0.025% or 0.05% colchicine, 49 were diploid with 20 chromosomes (Fig. 1b), and the other 47 were haploid with ten chromosomes. Of 53 plants from the calli treated with colchicine for 48 h, 29 were diploid plants. Calli treated for 72 h did not regenerate any haploid plants, with most being diploid plants except for one and four tetraploid plants with 40 chromosomes obtained from the two 72-h treatments of 0.025% and 0.05% colchicine, respectively. No significant difference in chromosome doubling was observed between these two colchicine levels (Table 1).

The haploid plants regenerated in this study all displayed a characteristic morphology (short, narrow leaves, reduced vigor, and no pollen shed). Under the same growing conditions, the doubled haploid plants

Table 1. Ploidy of plants regenerated from colchicine-treated haploid calli as determined from root tip squashes

Colchicine treatment		No. of plants regenerated			
Hours	Concentration	Total	Haploid	Diploid	Tetraploid
—	—	24	24	0	0
24	0.025%	48	23	25	0
	0.05%	48	24	24	0
Total		96	47	49	0
48	0.025%	22	8	14	0
	0.05%	31	16	15	0
Total		53	24	29	0
72	0.025%	31	0	30	1
	0.05%	32	0	28	4
Total		63	0	58	5

were generally more vigorous in appearance and grew more rapidly when compared with the haploid plants (Fig. 2). The doubled haploid plants from different treatments exhibited similar morphology. Most of them produced abundant, viable pollen. A common feature of many of the doubled haploid plants was the appearance of tassels with some female flowers. The ears of these plants could, however, still be self-pollinated if the silks emerged in time.

Most of the doubled haploid plants, 107 of 136, produced from 1 to 256 seed per ear after self-pollination. A few ears had almost normal seed set (Fig. 3). Among 29 doubled haploid plants which did not produce seed, 21 of the plants could not be pollinated due to asynchronous pollen shed and silk emergence, the lack of ear development, or to stunted growth. Eight other plants produced no seed even after being pollinated one or two times on successive days. The synchrony of pollen shed and silk emergence were the main factors which affected the seed production by the doubled haploid plants. As shown in Table 2, if the silks emerged for pollination 1–3 days later than the first pollen was shed, an average of more than 87 seeds per ear were set. If the pollination was started 4 days later than the first pollen was shed, the seed set was dramatically decreased to 39 seeds per ear. Most plants would not set seed if silk emergence was delayed 5 days or more after pollen shed began.

Five tetraploid plants were found among the plants regenerated after the two 72-h colchicine treatments. Of these five plants, two plants had terminal ears and three plants had good pollen shed, but due to late silk emergence, only two of the plants produced one seed each after self-pollination.

Anther-derived callus lines from two other hybrids, H99 × Pa91 and Pa91 × Fr16, were also treated with colchicine. Due to lower regenerability of the callus line from Pa91 × Fr16 and incomplete experiment design for the callus line from H99 × Pa91, the data from these two lines are not included. However, these experiments also showed that the longer the callus cultures were incubated

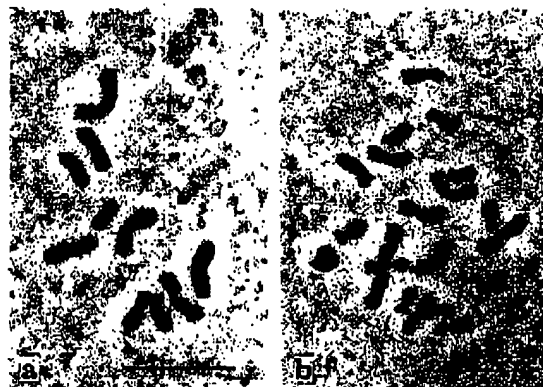


Fig. 1 a and b. Root tip chromosomes from a haploid cell with 10 chromosomes from a plant regenerated from untreated callus and b a diploid cell with 20 chromosomes from a plant regenerated from colchicine treated callus. Bar represents 10 μ m

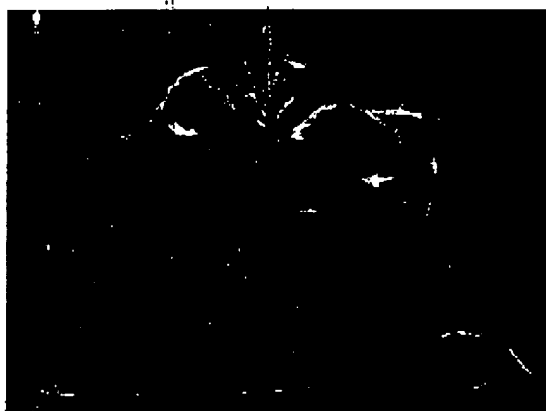


Fig. 2. Typical appearance of a doubled haploid plant (middle) from colchicine treated haploid callus, haploid plant (right) from colchicine-treated haploid callus, and haploid plant (left) from untreated callus. The pot diameters are 27.5 cm



Fig. 3. Mature ears resulting from self-pollination of some doubled haploid plants

Table 2. The relationship between the delay in silk emergence after the beginning of pollen shed and the average number of seed produced per ear from 33 randomly selected doubled haploid plants

Silk emergence delay (d)	No. of plants pollinated *	Average seeds per ear	No. of ears without seed
1	4	100.3	0
2	8	91.0	0
3	8	87.1	0
4	5	39.0	0
> 5	8	2.5	5

* One ear per plant was self-pollinated

in colchicine-containing medium, the more diploid plants were regenerated, and no diploid plants were regenerated from the control calli without colchicine treatment. These results then indicate that chromosome doubling of maize callus tissue by colchicine treatment is reproducible and is not genotype-specific.

Discussion

The present study shows that the colchicine treatment of the embryogenic haploid maize callus can be very effective for producing a large number of doubled haploid plants. By incubating embryogenic haploid calli on colchicine-containing medium, doubled haploid plants were produced at high frequencies. Since all the plants from untreated calli were haploids, the occurrence of doubled haploid plants must be due to the effect of colchicine. The method is rapid since it only required 6 months from colchicine treatment of calli to the harvest of seeds from the regenerated doubled haploid plant.

The results of this study suggest that the duration of colchicine treatment is important. The treatment of more than 48 h is necessary in order to get higher frequency of doubled haploids among the regenerated plants. If the treatment is 72 h, tetraploid plants could be produced, which may not be desirable. The two concentrations of colchicine used, 0.025% and 0.05%, did not show significant differences in their chromosome doubling efficiency.

There was no indication that ploidy chimeras were regenerated, since most of the doubled haploid plants produced seeds after self-pollination. The problem which caused the doubled haploid plants to not set seeds was mainly delayed silk emergence or the lack of ear formation, which are common phenomena among tissue culture-derived maize plants (Miao et al. 1978; Petolino and Jones 1986). The abnormal plants found among the regenerates were probably due to the tissue culture conditions rather than the colchicine treatment since the same abnormalities (stunted growth, terminal ear, the lack of normal ear) existed among the plants regenerated from untreated control calli. In practice, only vigorous plantlets should be selected before transplanting to greenhouse or field. This should reduce the frequency of abnormal plants.

The results show that colchicine treatment of embryogenic haploid callus can result in the production of

entire doubled haploid plants with high frequency, which produce fertile maize inbred lines within a short time at a high frequency, thus making the anther culture technique more useful to the plant breeder.

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References

- Chu CC, Wang CC, Sun CS, Hsu C, Yin KC, Chu CY (1975) Establishment of an efficient medium for anther culture of rice through comparative experiments on the nitrogen sources. *Sci Sin* 18:659-668
- Coc EH, Neuffer MG (1978) Embryo cells and their destinies in the corn plant. In: Subtelny S, Sumner IM (eds) *The clonal basis of development*. Academic Press, New York, pp 113-129
- Duncan DR, Williams ME, Zeiger BE, Widholm JM (1985) The production of callus capable of plant regeneration from immature embryos of numerous *Zea mays* genotypes. *Planta* 165:322-332
- Gamborg OL, Miller RA, Ojima K (1968) Nutrient requirements of suspension cultures of soybean root cells. *Exp Cell Res* 50:151-158
- Genovesi AD, Collins GB (1982) *In vitro* production of haploid plants of corn via anther culture. *Crop Sci* 22:1137-1144
- Ku MK, Cheng WC, Kuo LC, Kuan YL, An HP, Huang CH (1978) Induction factors and morpho-cytological characteristics of pollen-derived plants in maize (*Zea mays*). In: *Proceedings of symposium on plant tissue culture*. Science Press, Peking, pp 35-41
- Kuo CS, Lu WL, Kuo YL (1986) Corn (*Zea mays* L.): production of pure lines through anther culture. In: Bajaj YPS (ed) *Crops I (Biotechnology agriculture 2)*. Springer, Berlin Heidelberg New York, pp 168-180
- Miao SH, Kuo CS, Kuo YL, Sun AT, Ku SY, Lu WL, Wan YY, Chen ML, Wu MK, Hsu L (1978) Induction of pollen plants of maize and observations on their progeny. In: *Proceedings of symposium on plant tissue culture*. Science Press, Peking, pp 23-33
- Nitch C, Andersen S, Godard M, Neuffer MG, Sheridan WF (1982) Production of haploid plants of *Zea mays* and *Pennisetum* through androgenesis. In: Earle ED, Demarly Y (eds) *Variability in plants regenerated from tissue culture*. Praeger, New York, pp 69-91
- Petolino JP, Jones AM (1986) Anther culture of elite genotypes of maize. *Crop Sci* 26:1072-1074
- Petolino JP, Thompson SA (1987) Genetic analysis of anther culture response in maize. *Theor Appl Genet* 74:284-286
- Tsay HS, Miao SH, Widholm JM (1986) Factors affecting haploid plant regeneration from maize anther culture. *J Plant Physiol* 126:33-40
- Wu JL, Zhong LQ, Nong FH, Chen ML, Zhang HY, Zheng BL (1983) Selection of pure line of maize (*Zea mays*) by anther culture and observation on its hybrids. *Sci Sin* 26:725-733